# RSU JOURNAL OF BIOLOGY AND APPLIED SCIENCES ISSN: 2811 – 1451



# **ABOUT US**

Rivers State University Journal of Biology and Applied Science (RSUJBAS) publications is a quarterly, open access, international journal for all academic research in science discipline. Microbiology, botany, zoology, environmental biology, chemistry, physics, mathematics, computer science, biochemistry medical laboratory sciences and other applied science related areas. RSUJBAS is a platform set for elites to influence, contribute and communicate to the global environment through their various academic researches. We synergistically engage our noble effort to contribute to the knowledge development, discoveries and innovations in all fields of study. In RSUJBAS we publish research papers on current academic issues with standard scientific reviews. RSUJBAS publishes original research articles, review articles, case studies, short communications, survey report, comparative studies and many more.

### **Aims and Scope**

Rivers state University Journal of Biology and Applied Sciences aims to publish high quality papers that communicate fundamentals and contemporary discoveries both theoretical and practical. Most importantly RSUJBAS seeks to establish a platform for communicating emerging trends in various discipline such as Microbiology, Botany, Zoology, Environmental Biology, Chemistry, physics, Mathematics, Computer Sciences, Biochemistry, Medical Laboratory, Sciences, and other applied sciences related areas.

### **Description:**

- Area of concentration: All science academic disciplines
- Frequency of publishing: Quarterly
- Mode of publishing: both online and print publication
- Language of publication: English
- Double blinded Review Process
- Zero Level Plagiarism Tolerance

### Why Publish with us

Low Article Processing Charge (ACP) to promote the research work Easy and Rapid review process Instant publication upon acceptance Dedicated editorial and review team for fast review process RSUJBAS provides hard copies of publication every quarterly

# **EDITORIAL BOARD**

PROF. S.A. WEMEDO

Department of Microbiology Rivers State University

### PROF. C. K. WACHUKWU

Department of Medical Laboratory Science Rivers State University

### DR. (MRS) N.P. AKANI

Department of Microbiology River State University

### PROF.E.C. CHUKWU

Department of Plant Science and Biotechnology Rivers State University

### PROF. B.O. GREEN

Department of Plant Science and Biotechnology Rivers State University

### PROF. J.N. ONWUTEAKA

Department of Animal and Environmental Biology Rivers State University

### **DR. (MRS) A. P. UGBOMEH** Department of Animal and Environmental Biology Rives State University

**DR. (MRS) E. O. IBEGUDEM** Department of Medical Laboratory Science Rivers State University

**DR. F U. IGWE** Department of Biochemistry Rivers State University

**DR. V. I. E. ANIREH** Department of Computer Science Rivers State University

**DR. N. BOISA** Department of Chemistry

Rivers State University

### **DR. N. EBERE**

Department of Animal and Environmental Biology Rivers State University

**DR. D. O. NGEREBARA** Department of Geology

Rivers State University

**DR. D. MARTHIAS** Department of Computer Science Rivers State University

**PROF.G. C. AKANI.** Department of Animal AND Environmental Biology Rivers State University

**PROF.V.B. OMUBO-PEPPLE** Department of Physics Rivers State University

**DR. A.D. NWAOBURU** Department of Mathematics Rivers State University

**DR. A. R. C. AMAKIRI** Department of Physics Rivers State University

### DR. N. M. NAFO

Department of Mathematics Rivers State University

> All Correspondence to Prof Sam Wenedu (Editor -in -Chief) Department of Microbiology, Rivers State University

> > edictor.ibasya@yoo.com

Or

**OLUCHI DICKSON** 

Publication Manager dicksonoluchi87@gmail.com

# **CONSULTING EDITORS**

### Prof. F. O. Oroka

Department of Agronomy Delta State University, Abraka

### Naluba. N. Goddy (Ph.D.)

Department of Geography and Environmental Studies Faculty of Social Sciences, Ignatius Ajuru University of Education, Rumuolumeni, P.M.B.5047, Port Harcourt, Rivers State.

### Godpower- Echie, G.

Department of Integrated Science Ignatius Ajuru University of Education, Rumuolumeni, Port Harcourt.

## **GUIDELINE FOR MANUSCRIPTS**

Manuscripts should be typewritten on an A4sheet having B1.5=line spacing throughout the text. The margins should be 2B54cm (1 inch) in all sides and page number should be consecutively on the bottom of the page. The manuscript should be written in Times New Romans using '12' font size.

For original research paper, the manuscript should be arranged in the following order: Tittle page, Abstract, Keywords Introduction, Materials and Methods Results, Discussion, Acknowledgement, References, Tables with legends and supplementary materials

The tittle page should contain the title, the name(s) of the author(s), the name(s) and address (es) of the instruction(s) where the work was carried out, including a valid e-mail address from the corresponding author along with telephone numbers. The title of the manuscript should be specific and concise but sufficiently informative.

The Abstract should not exceed 250 words and it should contain brief summary of the findings including brief introduction, methodology, results, and conclusions,

The keywords should have a minimum of five and maximum of seven words.

The introduction should provide a clear statement of the problem and indicates aim of the study citing relevant literature to support background statements.

The Materials and Method should include the methods and methodology of the research.

The results should be presented in the form of tables of figures. It should be presented with clarity and precision. Statements used to present results should be written in the past tense. Detailed interpretation of data should not be included in the results but should be put into the Discussion section.

The Discussion should interpret the results clearly and concisely, and should integrate the research findings of this and past studies on the topic. Highlight the significant/unique findings of the research under conclusion.

The acknowledgment of people, grants or funds should be brief.

# **TABLE OF CONTENTS**

Creating fuel briquettes from discarded Coconut shells and peanut shells	1-12
as Renewable energy sources	
Abhon Adus & Egbe Ebiyeritei Wisdom	
Microbial Evaluation of Tap Water in Delta State University, Abraka	13-23

Microbial Evaluation of Tap Water in Delta State University, Abraka	
<u>Okolosi-Patani,</u> Omotejohwo Emily, <u>Akpo</u> , Christiana Orevaoghene &	
<u>Obiebi,</u> Nora	

### MICROBIAL EVALUATION OF TAP WATER IN DELTA STATE UNIVERSITY, ABRAKA <u>Okolosi-Patani</u>, Omotejohwo Emily Department of Pharmaceutical Microbiology and Biotechnology, Faculty of Pharmacy, Delta State University, Abraka, Nigeria.

### Akpo, Christiana Orevaoghene

Department of Medical Laboratory Science, Faculty of Science, Delta State University, *P.M.B 1* Abraka, Nigeria.
Email: <u>Cakpo@delsu.edu.ng</u>; <u>mummypure637@gmail.com</u>.

### Obiebi, Nora

Department of Pharmaceutical Microbiology and Biotechnology, Faculty of Pharmacy, Delta State University, Abraka Nigeria.

### ABSTRACT

Water is one of the most abundant resources on which life on earth depends; in some places, the availability of water is critical, limited, and renewable. The study is aimed at a microbial evaluation of the quality of tap water at Delta State University, Abraka. The pour plate method was used to determine the total heterotrophic count and thereafter, bacterial isolates were characterised using standard microbiological methods. The most probable number technique was employed in determining the total coliform count. Findings from the study revealed the heterotrophic count of the various water samples was between  $3.0 \times 10^4$  and  $3.4 \times 10^5$  cfu/ml. The coliform count in the presumptive test ranged from 9 to 240. This research also revealed the presence of bacteria species such as Pseudomonas *spp., Enterobacter spp.*, and *Aeromonas spp. Enterobacter spp.* occurred the most and accounted for 33.33%, while *Pseudomonas spp.* and *Aeromonas spp. accounted* for 16.67% each. *Escherichia coli* was absent in all the water samples analysed, but the presence of the isolated bacteria species in drinking water is of grave importance. The contamination of water samples may be a result of leaking underground pipes or inadequate sanitary conditions in the storage facilities. In order to improve the quality of grave importance.

Keywords: Tap-water, Microbial, Quality, Abraka.

### **INTRODUCTION**

Water is a unique commodity. It is the most abundant and essential resource among other natural resources useful to all living creatures (1). Water is the most indispensable need for the existence of all living things, for domestic and other purposes. Access to portable water is one of the many goals of the Sustainable Development Goals (SDG), which aim for environmental sustainability (2).

In mammals, water plays an essential role by absorbing a large amount of heat and then dissolving the waste products generated as a result of metabolism. Humans drink water, which provides for normal bodily and cell functions. A reduction in the daily intake of water causes a decrease in the efficiency of cells and other body activities. Humans also use water for agricultural, industrial, and recreational activities, in addition to other purposes (8, 9). Though all these needs are salient, water for human consumption and sanitation are regarded as having a significant social and economic impact on the health of the entire population.

Most of our water supplies are from surface water, which includes rivers, streams, lakes, oceans, seas, and other water bodies. In developing countries, water pollution and its shortage are prevalent (3). This may be due to poor domestic and personal hygiene and the accumulation of industrial and agricultural waste. As populations increase, the tendency for surface and underground water to be polluted becomes increasingly high. Basically, water-borne infectious diseases are transmitted primarily through water supplies contaminated with human and animal faecal matter. Some microorganisms isolated from water include *Escherichia coli, Staphylococcus sp., Bacillus sp., Streptococcus sp., Klebsiella sp., Pseudomonas sp., Flavobacterium sp., Enterobacter sp., Proteus sp., and Pseudomonas sp.* (4). Other human pathogenic organisms that pose a serious risk of

disease whenever found in drinking water include *Salmonella* spp., *Shigella spp.*, *Vibrio cholerae*, *Yersinia entercolitica*, *Campylobacter* spp., various viruses such as hepatitis E and A, rotaviruses, and parasites such as *Entamoeba histolytica* and *Giardia* spp. Previous findings reveal a high mortality rate from diarrheal-related diseases, and a greater percentage are children who are below the age of five (5).

It is important to state that water meant for consumption should not contain any microorganisms known to be pathogenic or any bacteria indicative of faecal pollution (6). Hence, it is absolutely necessary to test portable water to ensure it is safe. Evaluating the quality of portable water depends to a large extent on the examination of indicator organisms such as coliforms. One of such members of the faecal coliform group is *Escherichia coli*. *E. coli* is regarded as a specific indicator of faecal pollution in drinking water because of its prevalence in human and animal faeces as compared to other coliforms (7).

Disease outbreaks and economic losses could result from a shortage of water; hence, water is a necessity. Spores of bacterial and fungal origin may be present in polluted water. These spores may be obtained from the air, dead plants, animal sewage, and organic waste. Almost all microorganisms may be found in water, but bacteria appear to be the major water pollutants. It is important to know that dead, decaying organic matter acts as a substrate for some microorganisms (10). Bacteria are known to be helpful in the digestion of poisons from food and water. The presence of other species could cause various diseases in humans and other animals. It is also a known fact that water from wells, streams, rivers, boreholes, and even rainwater contains dissolved materials from the air as well as suspended dust intermixed with microorganisms (11).

At Delta State University, the topography of the land is flat, and the ground water level is between 6 and 8 meters. It is of utmost importance that drinking water from the source has to be tested and regularly monitored by the authorities to ensure that it is pure and germ-free. In this study, the bacteriological quality of the water being supplied to the DELSU community was microbiologically evaluated.

### EXPERIMENTAL

#### Study area

Delta State University, Abraka, is located in Delta State, Nigeria. It is a typical rural setting, although gradually transforming into a semi-urban area with the establishment of Delta State University. Abraka has an estimated population of 50,000. The major occupation of the indigenes is farming. Civil servants also abound in the community because of the presence of the University and public schools.

### Sample collection, transport, and storage

The water samples were collected from taps at different locations at Delta State University, while aseptic conditions were maintained during the collection of samples. Samples from the taps were taken after allowing the taps to run for about five minutes. Approximately 200 ml of water was collected from each tap location coded as A, B, C, D, E and F. The samples were kept in an ice pack to prevent any changes in the microbial flora of the samples during transport. The water samples were transported to the laboratory in a vertical position, maintaining a temperature of 1-4 °C with ice pack-enclosed conditions. Samples were analysed immediately after collection.

### Total heterotrophic bacteria count

The total bacterial count was carried out by the pour-plate method. In a sterile petri dish, one millilitre of each water sample was transferred aseptically and mixed with 19 ml of melted nutrient agar. The experiment was done in triplicate. The medium was allowed to solidify, and the plates were incubated at 37°C for 24 hours. The number of colonies in each plate was counted and the number of bacteria/ml of each sample, calculated.

### **Isolation and Identification of Bacteria**

Each water sample was subjected to serial dilution and 0.1 ml of the 10<sup>-5</sup> dilution was plated in MacConkey agar, blood agar, Mannitol salt agar, Salmonella-Shigella agar and Simmon Citrate agar. Incubation of the plates was done at37<sup>o</sup>C for 24 hours. Afterwards, the cultural appearance of the bacterial colonies were studied. This was followed by the Gram staining of the colonies. The biochemical tests performed on the bacterial isolates are indole, citrate utilization, oxidase, hydrogen sulphide, catalase, urease, MR-VP and carbohydrate fermentation.

### **ENUMERATION OF COLIFORM BACTERIA**

### Most probable number test

This is a 3 step test comprising of the Presumptive test, confirmed test and the completed test. The multiple tube fermentation technique was performed as a presumptive test for total coliform using a total of 9 tubes containing 5ml of MacConkey broth and inverted Durham tubes. To the first 3 tubes containing double-strength MacConkey broth, 10 ml of the original sample was added. Then to second set of 3 tubes containing single-strength MacConkey broth, 1 ml of the original sample was added. The last set of 3 tubes containing single-strength MacConkey broth, 1 ml of the original sample original sample added to them. All tubes were incubated at 37°C for 48 hours for the observation

of gas production. First reading was taken after 24 hours to record positive tubes, and the negative ones were incubated for another 24 hours.

**Confirmed test:** A loopful of inoculum was taken from tubes that were gas positive for the presumptive test and was inoculated into a tube containing 10 ml Brilliant green lactose broth medium. All tubes were incubated at 37°C for 48 hours for the observation of gas production.

### **Completed test (Fecal coliform test)**

At least 3 loopful of each confirmed positive tube were subcultured into EC (Escherichia coli) broth medium and then incubated at 44.5°C for 24 hours. Tubes showing any amount of gas production were considered as positive and the most probable number was recorded (the results were compared with the most probable number table) (12).

#### **RESULT AND DISCUSSION**

The heterotrophic count of the various water samples was between  $3.0 \times 10^4$  and  $3.4 \times 10^5$  cfu/ml (Table 1). A previous study reported the mean total bacterial counts of  $2.0 \times 10^4$  cfu/ml in borehole water (13). However, in a similar study surface water had a higher bacterial count compared to ground water (22). These counts clearly exceed the World Health Organization standard limit for drinking water (14) and therefore not acceptable. High bacterial counts is indicative of elevated level of the pollution of water, this could be attributed to both human and animal activities. Surface water can be contaminated by human waste such as sewage and bacteria along the flow paths of water (5). Other sources of contamination may include animal waste deposition, pasture and surface runoff. Of the six borehole water samples evaluated, two samples had zero coliform count (Table 2.0) thereby complying with the WHO standard for coliform in water (14).

Water	Heterotrophic Plate Count (Cfu /ml)
Samples	
А	$4.0 \ge 10^4$
В	$3.0 \ge 10^4$
С	$2.3 \text{ x} 10^5$
D	3.4 x 10 <sup>5</sup>
E	3.4 x 10 <sup>5</sup>
F	2.9 x 10 <sup>5</sup>

Table 1: Mean Heterotro	phic plate c	ount of the var	ious water samples
-------------------------	--------------	-----------------	--------------------

The highest coliform count in the presumptive test was 240 while the least was 9 (Table 2.0). These clearly exceeded the WHO standard. However, two out of the six samples had zero coliform count thereby complying with the WHO standard for drinking water. A similar study reported that coliform bacteria were present in all collected samples with MPN values ranging from 14 to 1600 (15) however findings from another study by Muhammad revealed a lower coliform count ranging from 14 to 36 counts (21). The incidence of coliform in water suggests the possible pollution by other infective microorganisms thus rendering such water unsafe for food processing also for consumption (16). The WHO guideline recommends further investigation for any water sample with high coliform counts (14).

Water sample	Number of tubes giving positive reactions				
	3 of 10ml	3 of 1ml	3 of 0.1ml	MPN/ml	
А	2	0	0	9	
В	3	0	0	23	
С	3	3	0	240	
D	0	0	0	0	
Е	3	3	0	240	
F	0	0	0	0	

### Table 2: Presumptive Test

This research also revealed the presence of three bacteria species namely *Pseudomonas spp*, Enterobacter spp and Aeromonas spp (Table 3.0). Enterobacter spp was most prevalent while *Pseudomonas* and *Aeromonas spp* had the least occurrence. This is clearly in contrast with a previous study done in Riyad, Saudi Arabia where the tap water was of a better quality (23) An earlier study revealed E. coli as most prevalent (61 %), P.aeruginosa 53%, Salmonella 25 % and S. aureus was least prevalent with 14 % (15). Laboratory investigations of water from River Ethiope in Abraka showed the presence of E. coli, S. aureus, Bacillus spp, S. epidermidis, Actinomyces and Enterobacter sp. (17). The occurrence of these bacterial species in drinking water is of grave importance. *Enterobacter spp* is known to be a fecal coliform which is indicative of faecal contamination of the water. Consumption of such water can result in health hazards (18). Numerous health risks have been associated with the consumption of faecal contaminated water. Immuno-compromised adults and children under the age of 5 are known to be at greater health risk if they consume faecal contaminated water (19). It has been documented that water borne diseases such as diarrhea, cholera dysentery typhoid, and polio amongst others are the major causes of death in underaged children (20) *Pseudomonas spp* on the other hand is capable of giving rise to urinary infections. The pollution of such water may be due to water the presence of sewage tanks around

the vicinity of the taps. Contamination can also result from underground leaking pipes meant to transport water from one location to another. Inadequate treatment of storage facilities can also give rise to storage tank harbouring some opportunistic pathogens such as *Pseudomonas spp* which can give rise to a number of infections in humans. Appropriate treatment processes must therefore be utilized in order to minimize health hazards resulting from the ingestion of such contaminated water (18).

Isolates	Α	В	С	D	Ε	F	Frequency of occurrence (%)
Pseudomonas	+	-	-	-	-	-	16.67
spp Enterobacter	-	+	+	-	-	-	33.33
spp Aeromonas spp	-	-	-	-	+	-	16.67

**Table 3: Frequency of Occurrence of Isolates from Water samples** 

### Conclusion

In conclusion, the presence of microorganisms of various species is in accordance with the fact that microorganisms are ubiquitous, occurring in air, water and also the environment. The water samples assessed had high heterotrophic counts and coliform counts, both of them clearly above the standard limit for drinking water. The presence of a feacal coliform indicates contamination of water with faecal matter. On the other hand *Pseudomonas spp* are capable of causing infection of the urinary tract. All of these microorganisms are harmful to human health. The contamination of water samples may be due to leaking underground pipes, inadequate sanitary conditions with the storage facilities and presence of sewage tanks around the vicinity of the taps and storage facilities.

In order to improve the quality of tap water in the community the storage facilities have to be maintained periodically, leaking pipes have to be fixed and human and animal waste have to be far from storage facilities and taps. In general, sanitary conditions should be improved in the campus.

### REFERENCES

- Abhishek, C., Pankaj, G., Ajit, V. and Tanu, J. (2017) Microbiological evaluation of drinking water sold by roadside vendors of Delhi, India. *Applied Water Science* **7**, 1635–1644.
- Ajayi AO, Adejumo TO (2011) Microbiological assessment and some physico-chemical properties of water sources in Akungba-Akoko, *Nigeria Journal of Toxicology and Environmental Health Sciences*, **3** (13), 342-346,
- Ajayi AO, Akonai KA (2005). Distribution pattern of enteric organisms in the Lagos Lagoon. *African Journal of Biomedical Research*, **8** (3), 163-168.
- Akpo CO and Mukoro J (2019). Bacteriological and Physiochemical Analysis of Water from River Ethiope, Abraka. *Sokoto Journal of Medical Laboratory Science*, **5**(1), 114 120.
- Allam RR, Uthappa CK, Nalini C, Udaragudi PR, Tadi GP, Murhekar MV (2015) An Outbreak of Cholera due to Contaminated Water, Medak District, Andhra Pradesh, India, 2013. Indian Journal of Community Medicine: Official Publication of Indian Association of Preventive Social Medicine, 40, 283–285.
- American Public Health Associated (APHA) (1998). Standard method for examination of water and waste water, 20th edn.American Water Work Associated (AWWA) Research Foundation. Denver.
- <u>Anyamene</u> NC, <u>Ojiagu</u> DK (2014). Bacteriological analysis of sachet water sold in Awka Metropolis, Nigeria, *International Journal of Agriculture and Biosciences*, **3** (3), 120-122.
- Diouf K, Tabatabai P, Rudolph J, Marx M (2014) Diarrhoea prevalence in children under five years of age in rural Burundi: An assessment of social and behavioural factors at the household level. *Global Health Action* 7, 24895.
- Edbert DC, Sandra AU, Ebere EC (2017). "Storage and its Effect on Chemical Quality Indicators in Sachet Water Brands Sold in Owerri Municipal," *Journal of World News of NaturalSciences*, **12**, 73–81,



- Egberongbe HO, Bello OO, Solate AT, and Sossou, MS. (2012) Microbiological evaluation of stream water for domestic use in rural areas: A case study of Ijebu North Local government, Ogun state, Nigeria. *Journal of Natural Sciences, Engineering and Technology*, **11**, 93-103
- Idibie OC, Onome EO, Oluwatoyin FI, Ambrose OE (2018). Comparative microbial analysis of borehole water and other sources of water in Benin Metropolis Edo State. *Journal of Environmental sciences and Public Health* **2**(4),232-242
- Kawther FA, Alwakeel SS (2007) Mineral and microbial content of Bottled and Tap water in Riyadh Saudi Arabia *Middle East Journal of Scientific Research*, **2**(3-4),151-156.
- Kuta G, Emigilati M, Hassan A, Ibrahim I (2014) "Domestic water sources and its health implication in Lapai Local Government area, Niger State, Nigeria," *Ethiopian Journal of Environmental Studies and Management*, 7 (6), 686-694
- Muhammad M, Samira S, Farrukh J, Faryal A (2013) "Assessment of Drinking Water Quality and its Impact on Residents Health in Bahawalpur City," *International Journal of Humanitiesand Social Science*, **3**, 114–128.
- Muhammad MN, Usoro KU, Ohworuka OG, Andrew N, Habib A (2015). Evaluation of drinking water quality at water distribution points in Nigerian Defence Academy. *International journal of Advanced research***3**(10), 1902 -1911
- Olasoji S, Oyewole N, Abiola B, Edokpayi J. (2019) Water Quality Assessment of Surface and Groundwater Sources Using a Water Quality Index Method: A Case Study of a Peri-Urban Town in Southwest, Nigeria. *Environments* [Internet] **6**(2), 23. Available from: http://dx.doi.org/10.3390/environments6020023
- Oyedeji O, Olutiola PO, Moninuola MA (2010). Microbiological quality of packaged drinking water brands marketed in Ibadan metropolis and Ile-Ife city in South Western Nigeria *African Journal of Microbiology Research* **4**(2), 96-102.
- Peter HR, George BJ (1989). *General Microbiology* 2nd edn.Water Treatment.*Time mirror/Mosby College publishing*. St Louis, USA
- Prasai T, Lekhak B, Joshi DR, Baral P (2007) Microbiological analysis of drinking water of Katmandu Valley. *Science World Journal*, **5**(5), 112–114.
- Prescott LM, Harley JP, Kleins DA (2008). *Microbiology*, 11th edn. Treatment of Water *McGram Hill Company*, New York.
- Shittu OB, Olaotan JO, Amusa TS (2008). Physicochemical and Bacteriological Analysis of water used for Drinking and Swimming Purposes in Abeokuta, Nigeria. *African Journal of Biomedical Research*, **2**, 285-290.



WHO (1997). *Guidelines for drinking water quality*, 2nd edn.Surveillance and control of community supplies, *World Health Organization*, Geneva.